



Red Load Taq Master / high yield

Master mix for direct gel loading

Cat. No.	Amount
PCR-106S-CSTM	1 ml (2x conc.)

For *in vitro* use only!

Shipping: shipped on blue ice

Storage Conditions: store at -20 °C

Additional Storage Conditions: avoid freeze/thaw cycles

Shelf Life: 12 months

Form: liquid

Concentration: 2x conc.

Description:

Red Load Taq Master / high yield contains an inherent red dye and allows the direct loading of the PCR reaction product onto the gel. It contains all reagents required for PCR (except template and primer) in a premixed 2x concentrated ready-to-use solution.

The Master Mix is recommended for use in routine PCR reactions. It is optimized for high efficiency and gives superior amplification results in a broad range of reaction conditions with most primer-template pairs. Note that the mix is based on a detergent containing buffer system not recommended for plate based PCR and automated pipetting.

The enzyme catalyzes the polymerization of nucleotides into duplex DNA in 5'→3' direction in the presence of magnesium. It also possesses a 5'→3' polymerization-dependent exonuclease replacement activity but lacks a 3'→5' exonuclease activity.

Kit contents:

2x Red Load Taq Master / high yield

master mix of thermostable DNA polymerase, dATP, dCTP, dGTP, dTTP, (NH₄)₂SO₄, MgCl₂, Tween-20, Nonidet P-40, red dye, gel loading buffer and stabilizers.

PCR grade water

Recommended 50 µl PCR assay:

25 µl	2x Taq Master Mix
0.2 - 1 µM	each Primer
2 - 50 ng	Template DNA
Fill up to 50 µl	PCR grade Water

Recommended cycling conditions:

Initial denaturation	94 °C	2 min	1x
Denaturation	94 °C	30 sec	30x
Annealing ¹⁾	45 - 68 °C	30 sec	30x
Elongation ²⁾	72 °C	30 sec - 3 min	30x
Final elongation	72 °C	2 min	1x

¹⁾The annealing temperature depends on the melting temperature of the primers used.

²⁾The elongation time depends on the length of the fragments to be

این فرآورده در شرکت تکاپوزیست از فرآورده عمده (Bulk) به شناسه PCR-106-CSTM تقسیم و بسته بندی شده است.

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amplified. A time of 1 min/kbp is recommended.

For optimal specificity and amplification an individual optimization of the recommended parameters may be necessary for each new template DNA and/or primer pair.

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